

情绪性摄食行为的神经环路调控机制

赵星琪, 柳丹阳, 周煜东*

(浙江大学脑科学与脑医学学院, 卫生部医学神经生物学重点实验室, 杭州 310058)

摘要: 摄食行为是机体维持个体生存和从事各种生命活动的保障, 它的调控十分复杂, 涉及到机体多个系统的协调参与, 其中中枢神经系统在摄食调控中起着关键性的作用。中枢神经系统不仅在因饥饿而摄食以维持能量稳态的过程中起关键作用, 还调控包括享乐进食在内的情绪性摄食行为。本文综述了中枢神经系统控制摄食行为的进展, 主要关注情绪性进食的中枢调控机制, 尤其是总结了中枢摄食、奖赏系统藉由各种投射路径和信号分子整合机体的营养、压力、情绪等信息进行摄食调控从而维持机体稳态的全过程。

关键词: 摄食; 情绪性摄食; 能量代谢; 奖赏; 食物成瘾; 压力应激

中图分类号: Q42; Q493.8 文献标志码: A

The central mechanism of emotional eating regulation

ZHAO Xing-Qi, LIU Dan-Yang, ZHOU Yu-Dong*

(NHC Key Laboratory of Medical Neurobiology, Department of Neurobiology,
Zhejiang University School of Medicine, Hangzhou 310058, China)

Abstract: Feeding behavior is key to supply energy for life activities and thus fundamental for survival. Feeding regulation is very complicated and requires the coordinated involvement of multiple systems of the body, among which the central nervous system(CNS) is of particular importance. The CNS not only controls hunger-related feeding behavior to maintain energy homeostasis, but also regulates emotional eating such as hedonic overeating. Here, we review the central mechanism of emotional eating regulation, with an emphasis on the interaction between the feeding and reward circuits via neural projections and signaling molecules that integrate the metabolic state and emotional salience to control emotional eating.

Key words: feeding; emotional eating; energy metabolism; reward; food addiction; stress

摄食行为 (feeding behavior) 是生物体为了满足生存所需要的能量, 在摄食动机的驱使下获取和消耗食物的行为, 是包括躲避天敌、繁衍后代在内的所有行为的基础。摄食的发生需要外周和中枢的协调参与。首先, 食物线索刺激视觉、嗅觉和味觉系统将信号传递至中枢神经系统 (central nervous system, CNS)^[1]; 然后, CNS 整合激素等生物活性物质的水平以及其他传入神经的输入信号, 综合评估个体的饥饿程度 (即饥饿感)、食物的营养价值^[2]、自身的代谢状态^[3] 和情绪状态以及进食风险等, 形成摄食动机即食欲, 进而调控摄食行为^[4]。

饥饿是产生摄食动机驱动摄食行为的重要因素, 因饥饿而摄食是维持机体能量稳态 (energy homeostasis) 的关键途径。饥饿能促进下丘脑摄食

中枢 (feeding center) 兴奋, 从而启动摄食行为。随着摄食的进行, 血糖上升, 饱腹感增加, 下丘脑饱食中枢 (satiety center) 相关神经元兴奋, 进而抑制进食。除此之外, 个体的进食动机也受其他因素调控。自身的情绪状态可以调控摄食行为, 比如正性的情绪反应能使动物在饱腹时继续进食, 这种摄食行为被称为情绪性摄食行为 (emotional eating)。情绪性摄食行为首先源于食物具有奖赏效应, 享用美味、高能量的食物带来的愉悦感会导致过量进食,

收稿日期: 2021-12-22; 修回日期: 2022-01-26

基金项目: 国家自然科学基金项目(82000801, 81770839)

*通信作者: E-mail: yudongzhou@zju.edu.cn; Tel: 0571-88208547

甚至食物成瘾。压力等造成的焦虑、抑郁等负性情绪状态同样可以影响摄食行为^[5],许多精神类疾病通常伴有进食行为异常。有意思的是,进食与代谢异常也会造成情绪障碍性疾病。减肥者在限量进食时会有情绪不稳定症状,包括肥胖在内的代谢类疾病增加焦虑症等疾病的患病风险^[6]。多种摄食障碍(eating disorders, EDS)疾病和焦虑症、重度抑郁症、强迫症等有40%~90%共患病率^[7]。这些摄食和情绪调节的交互作用提示,情绪调控机制在摄食中起着重要作用。现代社会的竞争加剧致使越来越多的人的心理处于亚健康状态,情绪异常引发EDS和代谢类疾病的发病率逐年攀升,因此情绪相关摄食行为越来越被人们关注。

随着神经示踪、脑成像、分子标记等技术的成熟,摄食行为的神经调控机制研究已得到了长足发展。本文将从摄食的神经环路机制入手,对目前CNS摄食中枢,尤其是情绪相关的核团调控摄食的研究进行综述。

1 稳态摄食行为的中枢机制

机体对能量的摄取和消耗在一定时间内通常处于相对平衡的状态。CNS对能量平衡的维持起着至关重要的作用。Kennedy^[8]和Gibbs等^[9]分别在1953和1973年提出了CNS存在影响摄食的“抑制信号”和“饱食信号”;之后的研究发现,这些信号主要产生于调控能量平衡和摄食行为的关键脑区——下丘脑。

下丘脑黑皮质素系统(melanocortin system, MC system)是由下丘脑中表达黑皮质受体(melanocortin receptors, MCRs)的神经元组成,主要位于下丘脑弓状核(arcuate nucleus, ARC),是摄食调控的关键系统^[10]。ARC中存在两类与摄食调控密切相关的细胞:一类是表达厌食肽前阿黑皮素原(pro-opiomelanocortin, POMC)的神经元;另一类神经元表达促进食欲的刺豚鼠相关蛋白(agouti-related protein, AgRP)和神经肽Y(neuropeptide Y, NPY)。长期激活ARC中的POMC神经元能减少食物的摄入量。用白喉毒素消融ARC中的POMC神经元会增加食物摄入,减少能量消耗,导致小鼠肥胖以及代谢和内分泌的紊乱^[11]。而利用光遗传学手段激活AgRP神经元能促进小鼠的摄食行为^[12],特异性杀死AgRP神经元则会导致其食欲下降,体重减轻,甚至死亡^[13]。上述两种神经元均表达MCRs。由POMC蛋白剪切而成的α-黑色素细胞刺激素

(α-melanocyte-stimulating hormone, α-MSH)可激活下一级神经元上的MCRs发挥抑制摄食作用^[14]。而AgRP则作为内源性的MCRs拮抗剂通过拮抗α-MSH的作用促进摄食行为的发生^[15-16]。现已发现MCRs中的MC4R和人类早发性肥胖相关,特异性敲除MC4R会导致小鼠高血糖、高胰岛素和肥胖等症状^[17]。

ARC位于第三脑室周围,此处血脑屏障薄弱,脑脊液中的营养物质以及激素等信号分子容易由此进入脑实质,因此ARC被认为是中枢感受外周信号的第一级核团。多种调控能量平衡的外周信号分子,如瘦素(leptin)、胰岛素(insulin)、胃饥饿素(ghrelin)和胆囊收缩素(cholecystokinin, CCK)等,均可作用于ARC。瘦素和胰岛素可分别作用于ARC上的相应受体,影响POMC神经元活动及NPY的合成和分泌,从而阻止食物摄入并调节体重^[18]。胃饥饿素进入体液循环而后透过血脑屏障进入ARC,激活AgRP神经元来释放促进食欲的NPY和AgRP。NPY、AgRP通过旁分泌和内分泌作用于下丘脑室旁核(hypothalamic paraventricular nucleus, PVN)等摄食相关脑区以及其他组织和器官,直接调控摄食行为。AgRP还可以通过分泌抑制性神经递质γ-氨基丁酸(γ-aminobutyric acid, GABA)与POMC神经元结合来抑制其活性,达到促进食欲的效果^[19]。

除了通过ARC感知体液内的摄食信号,CNS也可以通过神经信号传达饱腹感等信号。脑干的孤束核(nucleus of the solitary tract, NTS)为神经感知的主要核团,可接受肠胃迷走神经的信号进而影响摄食行为^[20]。NTS中也存在POMC神经元。和ARC不同,急性激活NTS中的POMC神经元会立即抑制进食,而消融这些POMC神经元后小鼠的体重和摄食却没有明显的变化^[11]。这些结果提示,在摄食行为调控过程中,以NTS为代表的神经通路对短时饱腹感信号更敏感,而以ARC为代表的体液通路则对长期能量稳态的调控更重要^[11,21]。

除了经典的MC system之外,CNS还存在多个脑区参与调控摄食行为。研究发现,下丘脑外侧区(lateral hypothalamic area, LH)、下丘脑腹内侧区(ventromedial hypothalamic nucleus, VMH)、PVN、中央杏仁核(central amygdaloid nucleus, CeA)、腹侧被盖区(ventral tegmental area, VTA)和中缝背核(dorsal raphe nucleus, DRN)等多个脑区均在摄食调控中发挥作用。LH中的许多神经元可以分泌食欲素(orexin)和黑色素浓缩激素(melanin concentrating hormone, MCH),

两者均是内源性促食欲分子。向大鼠的中枢系统中注射食欲素后可以促进其摄食行为^[22]。激活投射到VTA的食欲素能神经元可以抑制饱腹信号^[23]。MSH缺乏或MSH1受体被破坏的小鼠体型消瘦^[24]。VMH为经典的饱食中枢，可传递饱感信号。VMH中的一些神经元表达厌食性的脑源性神经营养因子(brain-derived neurotrophic factor, BDNF)，该因子或其受体表达降低会导致小鼠摄食增加，进而导致肥胖^[25]。此外，一些VMH细胞中含有葡萄糖感受器和脂肪酸感受器，可以感知机体血糖和脂肪酸水平并影响进食^[26-27]。

综上所述，当因能量消耗而偏离能量平衡，上述调控机制将共同作用来促进动物摄食行为的发生。当进食行为持续一段时间后，POMC神经元激活，AgRP神经元被抑制，体内激素等信号分子响应变化，进食停止，机体能量重新回归稳态。

2 正性情绪对摄食的调控

食物，尤其是富含糖和脂肪的食物，是一种天然的奖赏物。享用美味食物会有满足感和愉悦感，造成在没有额外能量需求的情况下过度进食，在这一过程中大脑的奖赏系统起了关键作用^[28]。和其他奖赏一样，食物奖赏也包括3种心理成分：由食物带来的快乐作用(hedonic impact)而喜欢(liking)、因食物具有激励显著性(incentive salience)而念想(wanting)^[29]以及建立线索和奖赏的预测关联(predictive associations)而学习(learning)。

中脑边缘多巴胺(dopamine, DA)系统(mesolimbic DA system)是调控食物奖赏效应的主要奖赏环路^[30]。DA奖赏系统包括VTA、伏隔核(nucleus accumbens, NAc)、内侧前额叶皮层(medial prefrontal cortex, mPFC)、海马(hippocampus, Hip)、基底外侧杏仁核(basolateral amygdala, BLA)和腹侧苍白球(ventral pallidum, VP)等。DA神经元主要分布于VTA^[31]，可口食物使中脑边缘奖赏系统DA含量迅速升高，参与编码“wanting”相关的行为动机，促进食物的摄入^[32]，DA合成障碍的小鼠摄食和饮水行为均明显减少^[33]。VTA脑区的DA神经元对NAc的GABA能抑制性神经元存在大量投射，是奖赏通路中的核心部分。投射到NAC的DA神经元损毁时，行动能力正常的小鼠获取食物的动机下降^[34]。VTA中的部分DA神经元也会投射到mPFC的谷氨酸能神经元，损毁mPFC不影响小鼠的摄食能力和稳态调节^[35]，但会影响味觉、嗅觉等从而影

响进食^[36-37]。VTA周期性地释放DA到NAc和Hip，响应奖赏预测线索等显著性信息，此过程对强化线索和奖赏的关联性学习至关重要^[38]。

DA奖赏系统和下丘脑摄食环路存在密切的神经连接，共同调控摄食行为。ARC的POMC和AgRP神经元与VTA神经元都存在直接的投射关系。ARC神经元释放的神经递质和神经肽可以作用于VTA脑区的神经元，动态调控摄食行为的完成。VTA的2种主要神经元——DA和GABA能神经元^[39]均表达MCRs^[40]，由POMC裂解而来的有活性的α-MSH既可以直接激活DA神经元促进DA释放，亦可以通过激活GABA神经元间接抑制DA神经元的功能。α-MSH对DA的作用效果主要以激活DA系统诱发奖赏效应为主^[41]，其对DA的抑制作用的生理功能还需进一步研究。AgRP神经元能通过释放GABA抑制DA神经元^[42]。下丘脑摄食环路激活DA奖赏通路的过程可总结如下：摄入食物后POMC神经元激活，其投射至VTA的轴突释放α-MSH与DA神经元的MCRs结合激活DA神经元，诱发奖赏效应^[41]；与此同时，处于抑制状态的AgRP神经元GABA释放减少，对DA神经元的抑制减弱，AgRP对DA神经元的抑制作用因此不足以抵消POMC神经元对DA神经元的激活效应^[43]，DA升高因而带来进食的满足感^[42]。

除DA外，大麻素、阿片类物质和5羟色胺(5-HT)等神经调质以及食欲素、瘦素等神经肽也参与食物的奖赏效应。在小鼠NAc处注射μ阿片受体激动剂会促进下丘脑分泌食欲素，改变其食物偏好，显著增加高脂肪食物的摄入^[44]。5-HT可以通过增强饱腹信号及其持续时间来抑制摄食^[45]，5-HT_{2C}受体激活POMC神经元，而5-HT_{1B}受体抑制NPY/AgRP神经元，达到抑制食欲的效果^[46]。因此，5-HT_{2C}/5-HT_{1B}受体缺乏的小鼠表现出食欲过盛和肥胖^[47]。急性给予食欲素和胃饥饿素可以激活大鼠VTA的DA神经元^[48]，引起奖赏效应。瘦素则可以直接或间接地影响VTA的DA神经元活性从而降低食物的奖赏效应，进而抑制摄食^[49]。

生理状态下的摄食行为是为了满足机体的能量需要，同时食物的奖赏性引发的愉悦感也会促进食物摄入。但当这种正性情绪带来的过量摄入可口食物的进食动机“劫持”奖赏系统后，会改变DA奖赏系统的功能、突触可塑性和DA的含量^[50]，尤其是NAc处的DA释放。奖赏系统对长期摄入的高能量食物产生的适应性改变，使机体在饱腹状态下

仍倾向于摄入食物, 不能及时停止过度的摄食行为, 从而引发肥胖, 甚至是在暴食、戒断、渴求等症间循环往复, 最终走向食物成瘾。此外, 奖赏系统过度激活促使个体在明知肥胖、健康状况受威胁的负面影响下仍选择继续进食, 最终发展为强迫性进食。

3 压力应激对摄食的影响

某些急性或慢性应激条件带来的压力, 如突发紧急事件、工作压力和社会污名等也会显著改变人们的摄食习惯和摄食类型。压力会破坏人认知层面的自我调节能力和情绪调节能力^[51], 为了缓解压力或转移注意力, 人们会选择增加或减少食物摄入。压力对摄食行为的促进或抑制作用尚无定论, 35%~60% 的人在压力状态下会吃更多的食物, 而另有 25%~40% 的人表示压力会抑制其进食能力^[52]。研究发现, 当面对心理压力时, 人们会更倾向于摄入高能量、高甜度的食物^[53]。动物实验也发现, 暴露于多种压力条件下的大鼠会摄入更多的猪油和蔗糖^[54]。压力导致的进食增加在肥胖人群中更为明显^[55], 这提示压力对摄食的影响也和人的代谢状态相关。压力引起不规律进食、改变食物偏好在能量代谢失衡的肥胖人群中可能更为显著^[56]。

压力可通过激活下丘脑-垂体-肾上腺轴(hypothalamic-pituitary-adrenal axis, HPA)影响摄食。下丘脑的 PVN 脑区作为 HPA 轴的起始点与多种压力应激相关。PVN 还可通过交感神经控制胃肠道分泌 CCK 并减少肠胃蠕动来抑制食欲^[57]。PVN 分泌的促肾上腺皮质激素释放因子(corticotropin-releasing factor, CRF)促进 POMC 衍生物——促肾上腺皮质激素(adrenocorticotropic hormone, ACTH)的释放, 最终导致皮质醇的合成增加。皮质醇可以直接促进摄食行为和脂肪沉积^[58], 还可以通过降低大脑对瘦素的敏感性^[59]以及调节 NPY 的分泌^[56]来促进摄食。皮质醇通过负反馈机制降低下丘脑 CRF 和 ACTH 的释放^[60], 同时增加瘦素和胃饥饿素的表达^[61], 调控能量稳态。长期激活 HPA 轴会改变瘦素、胃饥饿素、NPY 等多种摄食相关信号分子的表达, 改变葡萄糖代谢并引起瘦素抵抗^[62]。

压力引起的进食行为改变和 MC system 直接相关。POMC、NPY、AgRP 等神经肽以及 MCRs 响应压力变化调控摄食行为^[63]。小鼠连续 14 天在束缚器中经受 1 小时的慢性束缚压力应激造模后, ARC 的 POMC 神经元过度活化, 抑制 VTA 的 DA 神经元^[39], 进而减少摄食行为。连续 10 天经历

CD1 小鼠攻击的慢性社交挫败应激的小鼠, 其下丘脑 AgRP 和 NPY 的 mRNA 水平增加^[64], 并表现出对高脂高糖食物的偏爱^[65]。也有研究发现经历慢性社交挫败的小鼠食物摄入减少, 注射 MCRs 激动剂 MT II 会进一步降低进食, 而 MC4R 缺乏的小鼠表现出抵抗这种社交挫败应激的能力^[66]。

压力还可以激活奖赏系统, 增加 DA 的释放^[67-68]。人在嘈杂环境中进行高难度数学测试时引发的急性心理压力可增加奖赏寻求(reward seeking)行为, 促进高脂食物的消耗, 甚至引起强迫性进食^[69]。CRF、皮质醇和去甲肾上腺素等也可以作用于奖赏相关脑区使其敏感化, 进一步增强摄食动机。压力、奖赏和可口食物形成的正反馈循环可以缓解持续压力状态下的负性情绪^[56]。经常经历强压力还会改变 mPFC 以及边缘脑区的功能, 干扰情绪调控和行为控制, 并增加可口食物的摄入^[70]。

此外, 压力还可通过影响自身活动^[71]、睡眠质量^[72]和肠道微生物群^[73]等方式间接影响摄食行为。

4 负性情绪对摄食的影响

现代社会的高压状态致使越来越多人的心理处于亚健康状态。情绪低落、快感丧失以及过度强烈持续的担忧和恐惧等不良情绪状态往往伴随着食欲不振、进食困难等现象。焦虑症、重度抑郁症、创伤后应激障碍和 EDs 有 40%~90% 的共患病率, 情绪调节障碍是它们共同存在的病理机制。情绪障碍已被列为暴饮暴食症等 EDs 的关键预测因素^[74]。例如, EDs 患者情绪敏感多变且难以准确分辨自己和他人的情绪状态^[75]。明确负面情绪如何影响摄食行为对解决相关情绪性摄食障碍至关重要。

抑郁、恐惧和焦虑症患者和摄食障碍患者的情绪异常应该有其脑的结构和功能基础。研究表明, NAc、CeA、PFC、岛叶皮层(insular cortex, IC)、臂旁核(parabrachial nucleus, PBN)、腹侧纹状体(ventral striatum, VS) 和前扣带皮层(anterior cingulate cortex, ACC) 等脑区参与对外界刺激的评估和相应的情绪反应^[76]。虽然抑郁、焦虑等负面情绪和摄食行为的关联性已被广泛认可, 但它们的具体环路机制仍不明确。

流行病学研究发现, 高热量食物的消耗和焦虑症、抑郁症之间具有相关性^[77-78]。长时间(一个月)接触可口食物的大鼠在可口食物剥夺 24 小时后表现出明显的焦虑样行为, 小鼠 12 小时 10% 糖水 /12

小时糖水剥夺连续重复3~4周后也同样出现焦虑样行为^[79]。长期摄入富含碳水化合物食物的肥胖小鼠表现出焦虑样行为，同时PFC脑区的蛋白质降解程度增加^[80]。这些研究表明，长期摄入可口食物会诱发焦虑行为。近年的研究发现，IC-NAc环路调控焦虑心境下的摄食行为，激活该环路可减少小鼠的摄食^[81]。

长期摄入高能量食物还会诱发抑郁行为。大鼠用高糖饮食喂养7周后，在强迫性游泳实验和糖水偏好实验中均表现出抑郁样行为^[82]。用于抑郁症研究的先天无助大鼠模型也表现出了蔗糖偏好降低的抑郁样表型^[83]。目前认为，PFC等抑郁相关脑区体积减小、杏仁核和PFC脑区葡萄糖代谢异常以及HPA轴调节障碍在抑郁症的发生中发挥着重要作用^[84-85]。在负面情绪下，过度激活的杏仁核可能通过其下游中脑区域（如参与奖赏评估的NAc）提高可口食物的奖赏价值^[86]。

恐惧也可以改变摄食。恐惧引起的摄食行为变化与CeA、PBN和蓝斑（locus coeruleus, LC）三个脑区密切相关。激活CeA可诱发强烈的恐惧反应，CeA的过度激活也和创伤后应激障碍相关^[87]；损毁CeA可以消除多个物种的恐惧反应^[88]。CeA也在摄食调节中发挥作用：抑制CeA中表达前原孤啡肽（pronociceptin, PNOC）的细胞可以减少可口食物的摄入^[89]，而光激活CeA中表达蛋白激酶Cδ（PKCδ）的神经元则可通过局部释放GABA强烈抑制摄食^[90]。此外，胰岛素受体在CeA细胞上大量表达，在CeA处注射胰岛素可显著降低小鼠的摄食^[91]。PBN脑区可以被疼痛等伤害性刺激激活^[92]，主要传递恐惧信号至中枢；激活PBN也可以抑制摄食行为^[93]。PBN的降钙素基因相关肽（calcitonin gene-related peptide, CGRP）神经元会被新食物激活（新食物恐惧症），但在适应后，CGRP神经元会恢复正常摄食时的抑制状态^[92]。LC也参与恐惧引起的摄食抑制：LC神经元通过去甲肾上腺素和谷氨酸的共释放激活PBN神经元，以及通过诱导CeA到PBN抑制性投射的长时程抑制激活PBN神经元，介导“条件恐惧”引起的摄食抑制^[94]。以上研究表明，恐惧造成的CeA、PBN和LC相关神经元的激活是造成摄食抑制的神经机制。

5 摄食障碍与情绪异常

EDs是进食紊乱相关的一系列疾病的总称，神经性厌食症（anorexia nervosa, AN）、神经性贪食症

（bulimia nervosa, BN）以及暴饮暴食症（binge-eating disorder, BED）是常见的摄食障碍，主要表现为摄食习惯紊乱和对体重控制的偏执，会引发一系列生理和心理状态的紊乱。了解摄食相关疾病的病理特征对于疾病的预防和后续治疗至关重要，但现阶段对于这些疾病的认识仍十分有限。

值得注意的是，EDs也会造成情绪异常及相关精神症状。肥胖作为健康问题，经常伴随着抑郁和焦虑等情绪障碍，研究发现肥胖会显著增加抑郁症发作的风险^[95]。AN患者强迫性节制能量摄入，并对肥胖表现出强烈的恐惧样行为。BN患者以反复冲动性的进食伴随不适当的补偿行为为特征，临床发现患者常伴随精神分裂、重型抑郁等精神疾病^[96]。BED患者在暴饮暴食后会出现自责、焦虑等负性情绪。摄食障碍患者的临床治疗通常需要配合不同程度的心理治疗以达到改善或治愈的目的。

长期不健康的进食习惯会引发大脑发生适应性的改变，这是造成情绪异常以至于精神类疾病的原因。例如，长期摄入高热量食物会减少奖赏系统DA的分泌^[97]和降低DA受体的敏感性^[98]；利用脑成像技术观察到肥胖患者大脑纹状体Ⅱ型DA受体（D2R）的利用率^[99]以及合成DA的能力都有一定程度的下降；在过度摄食的小鼠模型^[100]和BED患者脑中^[101]均观察到了DA不足的现象；这些现象都符合成瘾的奖励缺乏理论，即由于DA神经元活动降低，相同价值的刺激带给成瘾者的愉悦感减少，需要代偿性的额外摄入来获得相同的愉悦感^[102]。DA神经元上存在瘦素、胰岛素等信号分子的受体，肥胖患者的瘦素、胰岛素抵抗现象也会间接影响DA神经元的活性，造成情绪的变化^[103]。EDs造成的神经系统的适应性改变还包括传递饱腹感的乙酰胆碱（acetylcholine, Ach）的释放延后^[104]，DA和Ach调控GABA-强啡肽输出使机体处于进食兴奋状态^[105]等。这一系列适应性变化导致机体需要暴露于更大的奖赏刺激才能达到和最初一致的愉悦水平^[106]，这与成瘾者的快感缺失一致。除此之外，暴食模型小鼠Hip-mPFC通路中存在BDNF和其受体原肌球蛋白受体激酶B（TRKB）表达异常、海马CA3区胰岛素受体表达减少、OFC处5-HT2_C受体上调等多种适应性变化^[107]，这些变化也可能会影响情绪相关通路的改变。

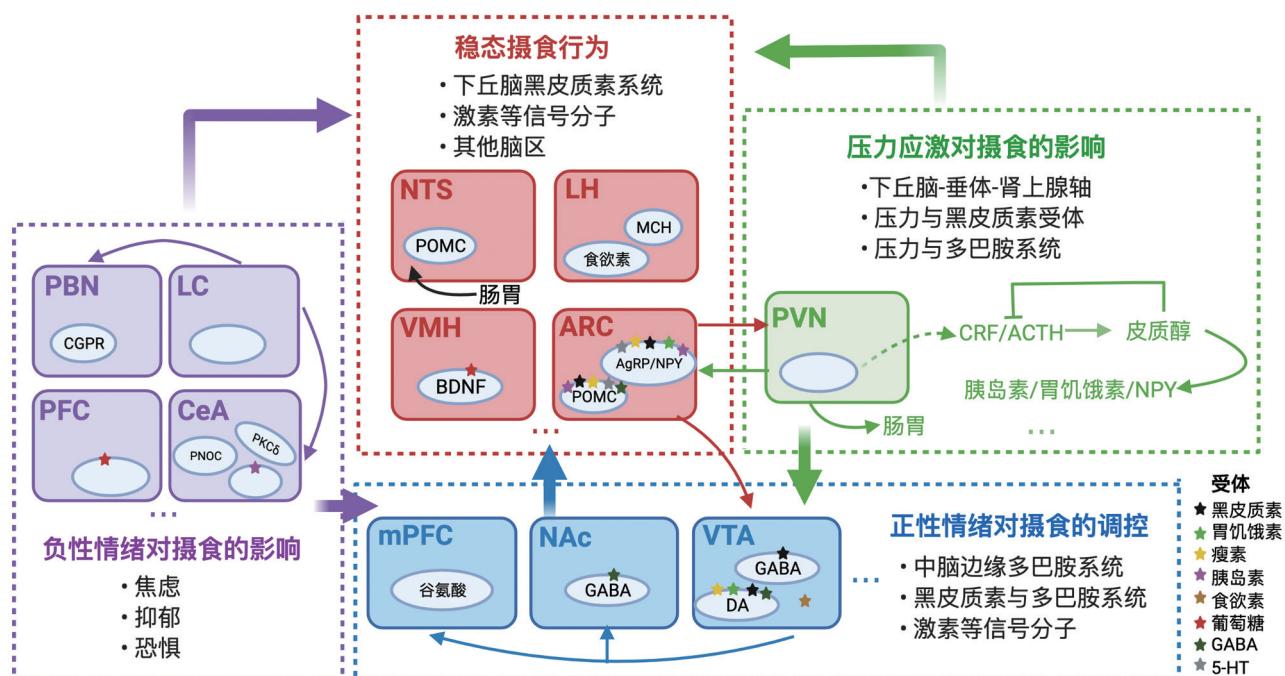
在大脑结构方面，近年来功能性磁共振脑成像（fMRI）研究发现了摄食障碍患者多个与情绪调控相关脑区的结构变化，为揭示情绪性摄食行为的脑机

制提供了结构基础^[108]。AN患者的脑区结构变化主要表现在杏仁核、ACC的体积减小，并且可在治疗后恢复正常。BN患者NAc脑区的体积减小，VS脑区的体积增加。急性BN患者的IC体积增加，康复后其脑区体积逐渐恢复正常。BN患者在面对意外奖赏以及奖赏缺失时，IC脑区的活性下降^[109]，这可能是由于EDs患者信息感受整合异常导致其摄食相关决策失误。NAc、VS、IC、杏仁核以及ACC等均与情绪调控密切相关，这些结果从结构上进一步提示了EDs会影响情绪体验。

6 总结和展望

摄食行为是机体维持个体生存、从事各种生命活动的保障，受到遗传、环境和社会心理等多因素交互作用的复杂调控。本文概述了情绪性进食的研究进展，总结了情绪体验和摄食行为交互作用的神经机制，特别是神经环路机制（图1）。这些研究进展得益于工具病毒的广泛使用，以及光遗传学、化学遗传学、在体电生理、荧光分子探针、功能性磁共振脑成像和钙成像等技术的发展。研究人员籍此可以精准定位、记录和操纵目标脑区特定目标神经元的活动，极大地方便了摄食行为的神经环路及机制研究。但目前摄食行为神经调控机制研究的瓶颈在于以下几方面。（1）我们对众多摄食脑区在摄食行为中的整合作用仍知之甚少。（2）许多研究局限在某些脑区可调控摄食行为的简单事实，但具体何种类型的神经元在其中发挥作用以及分子层面上的调控机制研究仍非常缺乏，而这些分子靶标可能是研发相关药物的重要突破点。荧光分子探针如多巴胺探针，可与钙成像、光遗传学技术结合，细胞特

情绪性摄食行为的神经环路调控机制



下丘脑ARC等核团(如NTS)藉由相关信号分子协调，参与调控稳态摄食行为。除此之外，自身的情绪状态也可以调控摄食行为。正性的情绪反应主要通过以VTA脑区为代表的多巴胺系统促进摄食，同时正性情绪也能通过黑皮质素系统及激素等信号分子参与摄食调控。压力应激对于摄食的影响主要通过HPA轴实现，同时压力也可作用于黑皮质素和多巴胺系统。焦虑、抑郁和恐惧等负性情绪可以通过不同的神经环路(如PBN、CeA)调控摄食行为。NTS，孤束核；LH，下丘脑外侧区；VMH，下丘脑腹内侧区；ARC，弓状核；POMC，促黑激素前阿黑皮素原；MCH，黑色素浓缩激素；AgRP，刺豚鼠相关蛋白；NPY，神经肽Y；BDNF，脑源性神经营养因子；PVN，下丘脑室旁核；CRF，促肾上腺皮质激素释放因子；ACTH，促肾上腺皮质激素；mPFC，内侧前额叶皮层；NAc，伏隔核；VTA，腹侧被盖区；GABA，γ-氨基丁酸；DA，多巴胺；PBN，臂旁核；LC，蓝斑；CeA，中央杏仁核；CGRP，降钙素基因相关肽；PNOC，前原孤啡肽；PKC δ ，蛋白激酶C δ ；5-HT，5-羟色胺。

图1 情绪性摄食行为的神经环路调控机制

异性地实现活体动物深部脑区中生物活性物质的动态记录^[110]。这类工具为将来更深入地研究食物的奖赏效应和成瘾机制以及相关药物靶点筛选提供了可能。(3) 噬齿动物模型实验的局限性导致心理和情绪等高级认知层面对摄食行为的调控作用的研究仍不够深入。未来利用多脑区和单细胞转录组结合功能研究技术，并建立灵长类情绪性摄食模型进行相关研究，将有助于更好地理解和治疗摄食障碍和代谢疾病。

参 考 文 献

- [1] Gutierrez R, Fonseca E, Simon SA. The neuroscience of sugars in taste, gut-reward, feeding circuits, and obesity. *Cell Mol Life Sci*, 2020, 77: 3469-502
- [2] Kaelberer MM, Buchanan KL, Klein ME, et al. A gut-brain neural circuit for nutrient sensory transduction. *Science*, 2018, 361: eaat5236
- [3] Carreiro AL, Dhillon J, Gordon S, et al. The macronutrients, appetite, and energy intake. *Annu Rev Nutr*, 2016, 36: 73-103
- [4] Prefit AB, Candea DM, Szentagotai-Tatar A. Emotion regulation across eating pathology: a meta-analysis. *Appetite*, 2019, 143: 104438
- [5] van Strien T. Causes of emotional eating and matched treatment of obesity. *Curr Diab Rep*, 2018, 18: 35
- [6] Sweeney P, Yang Y. Neural circuit mechanisms underlying emotional regulation of homeostatic feeding. *Trends Endocrinol Metab*, 2017, 28: 437-48
- [7] Stern SA, Bulik CM. Alternative frameworks for advancing the study of eating disorders. *Trends Neurosci*, 2020, 43: 951-9
- [8] Kennedy GC. The role of depot fat in the hypothalamic control of food intake in the rat. *Proc R Soc Lond B Biol Sci*, 1953, 140: 578-96
- [9] Gibbs J, Young RC, Smith GP. Cholecystokinin decreases food intake in rats. *J Comp Physiol Psychol*, 1973, 84: 488-95
- [10] Zhan C. POMC neurons: feeding, energy metabolism, and beyond. *Adv Exp Med Biol*, 2018, 1090: 17-29
- [11] Zhan C, Zhou J, Feng Q, et al. Acute and long-term suppression of feeding behavior by POMC neurons in the brainstem and hypothalamus, respectively. *J Neurosci*, 2013, 33: 3624-32
- [12] Aponte Y, Atasoy D, Sternson SM. AGRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. *Nat Neurosci*, 2011, 14: 351-5
- [13] Luquet S, Perez FA, Hnasko TS, et al. NPY/AgRP neurons are essential for feeding in adult mice but can be ablated in neonates. *Science*, 2005, 310: 683-5
- [14] Caruso V, Lagerstrom MC, Olszewski PK, et al. Synaptic changes induced by melanocortin signalling. *Nat Rev Neurosci*, 2014, 15: 98-110
- [15] Robinson SL, Thiele TE. The role of neuropeptide Y (NPY) in alcohol and drug abuse disorders. *Int Rev Neurobiol*, 2017, 136: 177-97
- [16] Cains S, Blomeley C, Kollo M, et al. AgRP neuron activity is required for alcohol-induced overeating. *Nat Commun*, 2017, 8: 14014
- [17] Huszar D, Lynch CA, Fairchild-Huntress V, et al. Targeted disruption of the melanocortin-4 receptor results in obesity in mice. *Cell*, 1997, 88: 131-41
- [18] Benite-Ribeiro SA, Rodrigues VAL, Machado MRF. Food intake in early life and epigenetic modifications of pro-opiomelanocortin expression in arcuate nucleus. *Mol Biol Rep*, 2021, 48: 3773-84
- [19] Cowley MA, Smart JL, Rubinstein M, et al. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature*, 2001, 411: 480-4
- [20] Akilan I, Sayar Atasoy N, Yavuz Y, et al. NTS catecholamine neurons mediate hypoglycemic hunger via medial hypothalamic feeding pathways. *Cell Metab*, 2020, 31: 313-26.e5
- [21] D'Agostino G, Lyons D, Cristiano C, et al. Nucleus of the solitary tract serotonin 5-HT2C receptors modulate food intake. *Cell Metab*, 2018, 28: 619-30.e5
- [22] Rani M, Kumar R, Krishan P. Role of orexins in the central and peripheral regulation of glucose homeostasis: evidences & mechanisms. *Neuropeptides*, 2018, 68: 1-6
- [23] Olney JJ, Navarro M, Thiele TE. The role of orexin signaling in the ventral tegmental area and central amygdala in modulating binge-like ethanol drinking behavior. *Alcohol Clin Exp Res*, 2017, 41: 551-61
- [24] Marsh DJ, Weingarth DT, Novi DE, et al. Melanin-concentrating hormone 1 receptor-deficient mice are lean, hyperactive, and hyperphagic and have altered metabolism. *Proc Natl Acad Sci U S A*, 2002, 99: 3240-5
- [25] Cheung CC, Kurrasch DM, Liang JK, et al. Genetic labeling of steroidogenic factor-1 (SF-1) neurons in mice reveals ventromedial nucleus of the hypothalamus (VMH) circuitry beginning at neurogenesis and development of a separate non-SF-1 neuronal cluster in the ventrolateral VMH. *J Comp Neurol*, 2013, 521: 1268-88
- [26] Viskaitis P, Irvine EE, Smith MA, et al. Modulation of SF1 neuron activity coordinately regulates both feeding behavior and associated emotional states. *Cell Rep*, 2017, 21: 3559-72
- [27] Le Foll C, Irani BG, Magnan C, et al. Characteristics and mechanisms of hypothalamic neuronal fatty acid sensing. *Am J Physiol Regul Integr Comp Physiol*, 2009, 297: R655-64
- [28] Rossi MA, Stuber GD. Overlapping brain circuits for homeostatic and hedonic feeding. *Cell Metab*, 2018, 27: 42-56
- [29] Morales I, Berridge KC. 'Liking' and 'wanting' in eating and food reward: brain mechanisms and clinical implications. *Physiol Behav*, 2020, 227: 113152
- [30] DiFeliceantonio AG, Small DM. Dopamine and diet-induced obesity. *Nat Neurosci*, 2019, 22: 1-2
- [31] Lutas A, Kucukdereli H, Alturkistani O, et al. State-specific gating of salient cues by midbrain dopaminergic input to basal amygdala. *Nat Neurosci*, 2019, 22: 1820-33

- [32] Yonemochi N, Ardianto C, Yang L, et al. Dopaminergic mechanisms in the lateral hypothalamus regulate feeding behavior in association with neuropeptides. *Biochem Biophys Res Commun*, 2019, 519: 547-52
- [33] Narayanan NS, Guarneri DJ, DiLeone RJ. Metabolic hormones, dopamine circuits, and feeding. *Front Neuroendocrinol*, 2010, 31: 104-12
- [34] Salamone JD, Wisniecki A, Carlson BB, et al. Nucleus accumbens dopamine depletions make animals highly sensitive to high fixed ratio requirements but do not impair primary food reinforcement. *Neuroscience*, 2001, 105: 863-70
- [35] Davidson TL, Chan K, Jarrard LE, et al. Contributions of the hippocampus and medial prefrontal cortex to energy and body weight regulation. *Hippocampus*, 2009, 19: 235-52
- [36] de Araujo IE, Rolls ET, Kringlebach ML, et al. Taste-olfactory convergence, and the representation of the pleasantness of flavour, in the human brain. *Eur J Neurosci*, 2003, 18: 2059-68
- [37] Rolls ET. Taste, olfactory and food texture reward processing in the brain and obesity. *Int J Obes (Lond)*, 2011, 35: 550-61
- [38] Monteleone AM, Castellini G, Volpe U, et al. Neuroendocrinology and brain imaging of reward in eating disorders: a possible key to the treatment of anorexia nervosa and bulimia nervosa. *Prog Neuropsychopharmacol Biol Psychiatry*, 2018, 80: 132-42
- [39] Qu N, He Y, Wang C, et al. A POMC-originated circuit regulates stress-induced hypophagia, depression, and anhedonia. *Mol Psychiatry*, 2020, 25: 1006-21
- [40] Alvaro JD, Taylor JR, Duman RS. Molecular and behavioral interactions between central melanocortins and cocaine. *J Pharmacol Exp Ther*, 2003, 304: 391-9
- [41] Roseberry AG, Stuhrman K, Dunigan AI. Regulation of the mesocorticolimbic and mesostriatal dopamine systems by alpha-melanocyte stimulating hormone and agouti-related protein. *Neurosci Biobehav Rev*, 2015, 56: 15-25
- [42] Dietrich MO, Bober J, Ferreira JG, et al. AgRP neurons regulate development of dopamine neuronal plasticity and nonfood-associated behaviors. *Nat Neurosci*, 2012, 15: 1108-10
- [43] Betley JN, Xu S, Cao ZFH, et al. Neurons for hunger and thirst transmit a negative-valence teaching signal. *Nature*, 2015, 521: 180-5
- [44] Zheng H, Patterson LM, Berthoud HR. Orexin signaling in the ventral tegmental area is required for high-fat appetite induced by opioid stimulation of the nucleus accumbens. *J Neurosci*, 2007, 27: 11075-82
- [45] Bacque-Cazeneuve J, Bharatiya R, Barriere G, et al. Serotonin in animal cognition and behavior. *Int J Mol Sci*, 2020, 21: 1649
- [46] Sohn JW, Elmquist JK, Williams KW. Neuronal circuits that regulate feeding behavior and metabolism. *Trends Neurosci*, 2013, 36: 504-12
- [47] Bouwknecht JA, van der Gugten J, Hijzen TH, et al. Male and female 5-HT(1B) receptor knockout mice have higher body weights than wildtypes. *Physiol Behav*, 2001, 74: 507-16
- [48] Cone JJ, McCutcheon JE, Roitman MF. Ghrelin acts as an interface between physiological state and phasic dopamine signaling. *J Neurosci*, 2014, 34: 4905-13
- [49] Domingos AI, Vaynshteyn J, Voss HU, et al. Leptin regulates the reward value of nutrient. *Nat Neurosci*, 2011, 14: 1562-8
- [50] Cocciarello R, Maccarrone M. Hedonic eating and the “delicious circle”: from lipid-derived mediators to brain dopamine and back. *Front Neurosci*, 2018, 12: 271
- [51] Raio CM, Oreduru TA, Palazzolo L, et al. Cognitive emotion regulation fails the stress test. *Proc Natl Acad Sci U S A*, 2013, 110: 15139-44
- [52] Packard AE, Ghosal S, Herman JP, et al. Chronic variable stress improves glucose tolerance in rats with sucrose-induced prediabetes. *Psychoneuroendocrinology*, 2014, 47: 178-88
- [53] Gibson EL. The psychobiology of comfort eating: implications for neuropharmacological interventions. *Behav Pharmacol*, 2012, 23: 442-60
- [54] Pecoraro N, Reyes F, Gomez F, et al. Chronic stress promotes palatable feeding, which reduces signs of stress: feedforward and feedback effects of chronic stress. *Endocrinology*, 2004, 145: 3754-62
- [55] Laitinen J, Ek E, Sovio U. Stress-related eating and drinking behavior and body mass index and predictors of this behavior. *Prev Med*, 2002, 34: 29-39
- [56] Sinha R, Jastreboff AM. Stress as a common risk factor for obesity and addiction. *Biol Psychiatry*, 2013, 73: 827-35
- [57] Ulrich-Lai YM, Fulton S, Wilson M, et al. Stress exposure, food intake and emotional state. *Stress*, 2015, 18: 381-99
- [58] Tomiyama AJ. Stress and obesity. *Annu Rev Psychol*, 2019, 70: 703-18
- [59] Jequier E. Leptin signaling, adiposity, and energy balance. *Ann N Y Acad Sci*, 2002, 967: 379-88
- [60] Dallman MF, Pecoraro NC, la Fleur SE. Chronic stress and comfort foods: self-medication and abdominal obesity. *Brain Behav Immun*, 2005, 19: 275-80
- [61] Chuang JC, Zigman JM. Ghrelin's roles in stress, mood, and anxiety regulation. *Int J Pept*, 2010, 2010: 460549
- [62] Milaneschi Y, Simmons WK, van Rossum EFC, et al. Depression and obesity: evidence of shared biological mechanisms. *Mol Psychiatry*, 2019, 24: 18-33
- [63] Maniam J, Morris MJ. The link between stress and feeding behaviour. *Neuropharmacology*, 2012, 63: 97-110
- [64] Patterson ZR, Khazall R, MacKay H, et al. Central ghrelin signaling mediates the metabolic response of C57BL/6 male mice to chronic social defeat stress. *Endocrinology*, 2013, 154: 1080-91
- [65] Moore CJ, Lowe J, Michopoulos V, et al. Small changes in meal patterns lead to significant changes in total caloric intake. Effects of diet and social status on food intake in female rhesus monkeys. *Appetite*, 2013, 62: 60-9
- [66] Chuang JC, Krishnan V, Yu HG, et al. A β 3-adrenergic-

- leptin-melanocortin circuit regulates behavioral and metabolic changes induced by chronic stress. *Biol Psychiatry*, 2010, 67: 1075-82
- [67] Dallman MF. Stress-induced obesity and the emotional nervous system. *Trends Endocrinol Metab*, 2010, 21: 159-65.
- [68] Wand GS, Oswald LM, McCaul ME, et al. Association of amphetamine-induced striatal dopamine release and cortisol responses to psychological stress. *Neuropsychopharmacology*, 2007, 32: 2310-20
- [69] Lemmens SG, Rutters F, Born JM, et al. Stress augments food ‘wanting’ and energy intake in visceral overweight subjects in the absence of hunger. *Physiol Behav*, 2011, 103: 157-63
- [70] Sinha R. Chronic stress, drug use, and vulnerability to addiction. *Ann N Y Acad Sci*, 2008, 1141: 105-30
- [71] Kim JS, Han SY, Iremonger KJ. Stress experience and hormone feedback tune distinct components of hypothalamic CRH neuron activity. *Nat Commun*, 2019, 10: 5696
- [72] Mu P, Huang YH. Cholinergic system in sleep regulation of emotion and motivation. *Pharmacol Res*, 2019, 143: 113-8
- [73] Kelly JR, Keane VO, Cryan JF, et al. Mood and microbes: gut to brain communication in depression. *Gastroenterol Clin North Am*, 2019, 48: 389-405
- [74] Hudson JI, Hiripi E, Pope HG Jr, et al. The prevalence and correlates of eating disorders in the National Comorbidity Survey Replication. *Biol Psychiatry*, 2007, 61: 348-58
- [75] Garcia SC, Mikhail ME, Keel PK, et al. Increased rates of eating disorders and their symptoms in women with major depressive disorder and anxiety disorders. *Int J Eat Disord*, 2020, 53: 1844-54
- [76] Donofry SD, Roecklein KA, Wildes JE, et al. Alterations in emotion generation and regulation neurocircuitry in depression and eating disorders: a comparative review of structural and functional neuroimaging studies. *Neurosci Biobehav Rev*, 2016, 68: 911-27
- [77] Quirk SE, Williams LJ, O’Neil A, et al. The association between diet quality, dietary patterns and depression in adults: a systematic review. *BMC Psychiatry*, 2013, 13: 175
- [78] Sanchez-Villegas A, Toledo E, de Irala J, et al. Fast-food and commercial baked goods consumption and the risk of depression. *Public Health Nutr*, 2012, 15: 424-32
- [79] Avena NM, Rada P, Hoebel BG. Evidence for sugar addiction: behavioral and neurochemical effects of intermittent, excessive sugar intake. *Neurosci Biobehav Rev*, 2008, 32: 20-39
- [80] Souza CG, Moreira JD, Siqueira IR, et al. Highly palatable diet consumption increases protein oxidation in rat frontal cortex and anxiety-like behavior. *Life Sci*, 2007, 81: 198-203
- [81] Gehrlach DA, Dolensek N, Klein AS, et al. Aversive state processing in the posterior insular cortex. *Nat Neurosci*, 2019, 22: 1424-37
- [82] Iemolo A, Valenza M, Tozier L, et al. Withdrawal from chronic, intermittent access to a highly palatable food induces depressive-like behavior in compulsive eating rats. *Behav Pharmacol*, 2012, 23: 593-602
- [83] Vollmayr B, Bachteler D, Vengeliene V, et al. Rats with congenital learned helplessness respond less to sucrose but show no deficits in activity or learning. *Behav Brain Res*, 2004, 150: 217-21
- [84] Koolschijn PC, van Haren NE, Lensvelt-Mulders GJ, et al. Brain volume abnormalities in major depressive disorder: a meta-analysis of magnetic resonance imaging studies. *Hum Brain Mapp*, 2009, 30: 3719-35
- [85] Jacques A, Chaaya N, Beecher K, et al. The impact of sugar consumption on stress driven, emotional and addictive behaviors. *Neurosci Biobehav Rev*, 2019, 103: 178-99
- [86] Bohon C, Stice E. Negative affect and neural response to palatable food intake in bulimia nervosa. *Appetite*, 2012, 58: 964-70
- [87] Bryant RA, Felmingham K, Kemp A, et al. Amygdala and ventral anterior cingulate activation predicts treatment response to cognitive behaviour therapy for post-traumatic stress disorder. *Psychol Med*, 2008, 38: 555-61
- [88] Janak PH, Tye KM. From circuits to behaviour in the amygdala. *Nature*, 2015, 517: 284-92
- [89] Hardaway JA, Halladay LR, Mazzone CM, et al. Central amygdala prepronociceptin-expressing neurons mediate palatable food consumption and reward. *Neuron*, 2019, 102: 1088
- [90] Uribe-Cerda S, Morselli E, Perez-Leighton C. Updates on the neurobiology of food reward and their relation to the obesogenic environment. *Curr Opin Endocrinol Diabetes Obes*, 2018, 25: 292-7
- [91] Mitchell CS, Begg DP. The regulation of food intake by insulin in the central nervous system. *J Neuroendocrinol*, 2021: e12952
- [92] Campos CA, Bowen AJ, Roman CW, et al. Encoding of danger by parabrachial CGRP neurons. *Nature*, 2018, 555: 617-22
- [93] Palmiter RD. The parabrachial nucleus: CGRP neurons function as a general alarm. *Trends Neurosci*, 2018, 41: 280-93
- [94] Yang B, Sanches-Padilla J, Kondapalli J, et al. Locus coeruleus anchors a trisynaptic circuit controlling fear-induced suppression of feeding. *Neuron*, 2021, 109: 823-38.e6
- [95] Luppino FS, de Wit LM, Bouvy PF, et al. Overweight, obesity, and depression: a systematic review and meta-analysis of longitudinal studies. *Arch Gen Psychiatry*, 2010, 67: 220-9
- [96] Wade TD. Recent research on bulimia nervosa. *Psychiatr Clin North Am*, 2019, 42: 21-32
- [97] Avena NM, Rada P, Hoebel BG. Sugar and fat bingeing have notable differences in addictive-like behavior. *J Nutr*, 2009, 139: 623-8
- [98] Bello NT, Lucas LR, Hajnal A. Repeated sucrose access influences dopamine D2 receptor density in the striatum. *Neuroreport*, 2002, 13: 1575-8
- [99] de Weijer BA, van de Giessen E, van Amelsvoort TA, et al.

- al. Lower striatal dopamine D2/3 receptor availability in obese compared with non-obese subjects. *EJNMMI Res*, 2011, 1: 37
- [100] Johnson PM, Kenny PJ. Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nat Neurosci*, 2010, 13: 635-41
- [101] Cameron JD, Chaput JP, Sjodin AM, et al. Brain on fire: incentive salience, hedonic hot spots, dopamine, obesity, and other hunger games. *Annu Rev Nutr*, 2017, 37: 183-205
- [102] Gondre-Lewis MC, Bassey R, Blum K. Pre-clinical models of reward deficiency syndrome: a behavioral octopus. *Neurosci Biobehav Rev*, 2020, 115: 164-88
- [103] Thompson JL, Borgland SL. Presynaptic leptin action suppresses excitatory synaptic transmission onto ventral tegmental area dopamine neurons. *Biol Psychiatry*, 2013, 73: 860-8
- [104] Avena NM, Bocarsly ME, Rada P, et al. After daily bingeing on a sucrose solution, food deprivation induces anxiety and accumbens dopamine/acetylcholine imbalance. *Physiol Behav*, 2008, 94: 309-15
- [105] Hoebel BG, Avena NM, Bocarsly ME, et al. Natural addiction: a behavioral and circuit model based on sugar addiction in rats. *J Addict Med*, 2009, 3: 33-41
- [106] Ifland JR, Preuss HG, Marcus MT, et al. Refined food addiction: a classic substance use disorder. *Med Hypotheses*, 2009, 72: 518-26
- [107] Chawla A, Cordner ZA, Boersma G, et al. Cognitive impairment and gene expression alterations in a rodent model of binge eating disorder. *Physiol Behav*, 2017, 180: 78-90
- [108] Wonderlich JA, Bershad M, Steinglass JE. Exploring neural mechanisms related to cognitive control, reward, and affect in eating disorders: a narrative review of fMRI studies. *Neuropsychiatr Dis Treat*, 2021, 17: 2053-62
- [109] Frank GK, Reynolds JR, Shott ME, et al. Altered temporal difference learning in bulimia nervosa. *Biol Psychiatry*, 2011, 70: 728-35
- [110] Sun F, Zeng J, Jing M, et al. A genetically encoded fluorescent sensor enables rapid and specific detection of dopamine in flies, fish, and mice. *Cell*, 2018, 174: 481-96. e19